

Research article – Review type.

Systematic review of genetic determinant and prevalence of resistance in bovine mastitis pathogens in Algeria

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Abstract

Mastitis is one of the most problematic and costly production diseases of dairy cattle. It is frequently treated with broad-spectrum antimicrobials. The aim of this paper is to document the current situation of antimicrobial resistance identified in the main pathogens causing bovine mastitis in Algeria. Genetic determinants and prevalence of resistance to different antimicrobials are discussed. Only studies focusing on bovine mastitic milk samples that investigated antibiotic-resistant bacteria were selected and just 7 articles met the inclusion criteria and were included in this systematic review. Data were narratively synthesized and analyzed.

The most commonly isolated bacteria were *Staphylococcus aureus* and *Escherichia coli*. The findings indicate signs of antibiotic misuse in Algeria, coupled with the rapid dissemination of resistant bacterial strains and inadequate surveillance, contributing to the problem. The resistance profile of the agents studied in Algeria showed alarmingly high levels of resistance to widely used antimicrobial drugs for the treatment of mastitis. Several genetic determinants were detected (*bla CTX-M*, *tetA*, *bla TEM-1*, *mecA*, *tetM*, *blaZ*).

Without critical measures to stop the excessive use of antibiotics, the prevalence of antibiotic-resistant bacteria could rise posing major risks to both animal and human health. Therefore, routine antimicrobial susceptibility testing before treatment is crucial for selecting effective antibiotics and preventing resistance development.

Keywords: Antibiotic therapy, Bovine mastitis, Resistance genes, Algeria, Systematic review

Introduction

Mastitis is one of the most prevalent diseases in dairy cows, which has an impact on the udder's health as well as the quality of milk yielded (Tommasoni *et al.*, 2023). Treatment for mastitis often consists of using broad-spectrum antibiotics, which are known to partially enhance resistance evolution, without any prior knowledge of cause-related agents. This increases the selective pressure on potentially present pathogens and is considered a potential human health risk (Rana *et al.*, 2022). *Methicillin-resistant Staphylococcus aureus* (MRSA) and multidrug resistant Gram-negative bacteria are considered a key challenge in mastitis treatment (Naranjo-Lucena and Slowey, 2023).

Mastitis control requires a comprehensive strategy, including herd disease prevention, antimicrobial stewardship, and targeted antibiotic therapy based on pathogen identification and resistance patterns. This is because there are many subclinical infections, withdrawal periods, and antimicrobial resistance (Ruegg, 2021).

Using the "One Health Approach" is essential to addressing the challenges caused by antibiotic resistance in the management of mastitis. This approach recognizes the interdependency of human, animal, and environmental health, striving to enhance comprehension and control of antimicrobial resistance concerning mastitis and other food. Key steps in this approach involve monitoring the prevalence of antimicrobial resistance in mastitis pathogens and implementing effective control measures (Sipahi *et al.*, 2023).

Thus, this review aims to document the current situation of antimicrobial resistance identified in the main pathogens causing bovine mastitis in Algeria. Genetic determinants and prevalence of resistance to different antimicrobials are discussed.

Methodology

An exhaustive search was undertaken across various databases, including Google Scholar, PubMed, Scopus and ScienceDirect, to identify relevant studies published over time. The search strategy incorporated specific headings and keywords such as "Antibiotic therapy," "bovine mastitis," "Algeria," and "Resistance genes." The review methodology involved a meticulous analysis of each identified paper, with a focus on extracting pertinent information from all sections to compile a comprehensive synthesis. To ensure data integrity, duplicate papers were systematically removed, and the remaining data underwent rigorous screening. Irrelevant works were excluded, and full-text documents were subsequently screened. The inclusion criteria were defined to encompass articles conducted in Algeria that specifically addressed antibiotics resistance and resistance genes isolated from agents responsible for bovine mastitis. Exclusion

criteria were applied to studies that did not utilize bovine milk mastitis samples, did not discuss resistance genes, or were not conducted in Algeria, Table 1.

Results

The systematic review identified a limited pool of literature meeting our predefined inclusion criteria. Only seven articles were deemed eligible for inclusion in our analysis. It is important to note that these studies were conducted in different parts of Algeria and at different periods and they used different methods to study and analyse. Even so, the prevalence of the bacteria may have changed over time and results may not be directly comparable. All of the eligible studies described the occurrence of antimicrobial resistance while all the studies did not provide information on farms types. The details of the studies regarding the countries; author name; year; Sampling date; reported prevalence of bacteria are given in Table 1.

Table 1. Prevalence of *S. aureus* and *E. coli* in Algerian dairy cows, highlighting a high occurrence of both pathogens, with significant antibiotic resistance patterns observed.

Region	Sampling date	Number of milk samples	Agent	Number of strains	References
Alger	Spring 2017 to Winter 2019	360	<i>Escherichia coli</i>	26,9 %	(Ghallache <i>et al.</i> , 2021)
Bordj Bou Arreridj	December 2016 to November 2018	200	<i>Escherichia coli</i>	26 %	(Tahar <i>et al.</i> , 2020)
Oran	February 2011 to February 2012	213	<i>Staphylococcus aureus</i>	38,9 %	(Benhamed and Kihal, 2013)
Tizi-Ouzou, Boumerdes, Khenchela, Blida	/	218	<i>Staphylococcus aureus</i>	29,8 %	(Akkou <i>et al.</i> , 2016)
Alger	2011 to 2013	/	<i>Staphylococci</i>	21 isolates	(Saidi <i>et al.</i> , 2015)
Batna, Constantine, Oum El Bouaghi, Setif	August 2015 to March 2016	167	<i>Staphylococcus aureus</i>	63,5%	(Zaatout <i>et al.</i> , 2019)
Tiaret	23 February to 23 June 2021	/	<i>Staphylococcus aureus</i>	62 isolates	(Bouzidi <i>et al.</i> , 2023)

Discussion

Mastitis can be categorized into clinical and subclinical mastitis based on different features. Clinical mastitis is characterized by visible abnormalities in the milk, such as flakes, clots, or watery secretions, as well as swollen, hot, and painful quarters. In acute cases, general signs like hyperthermia, anorexia, and depression can also be present (Cobirka *et al.*, 2020). On the other hand, subclinical mastitis is not easily diagnosed due to the lack of evident signs in the milk or the affected animals. Its main symptoms are associated with an increased somatic cell count and decreased milk production (Cobirka *et al.*, 2020).

Subclinical mastitis, as evidenced by studies in Algeria has been shown to have a high prevalence, with rates of 66.4% (Ghallache *et al.*, 2021), 52.12% (Benhamed and Kihal, 2013b) and 40% (Zaatout *et al.*, 2019) based on California Mastitis Test (CMT) results. This is concerning due to the potential for a longer duration of subclinical mastitis compared to clinical mastitis, as highlighted by Cobirka *et al.* (2020). The prolonged duration of subclinical mastitis can facilitate the transmission of pathogens within the herd. Therefore, these findings emphasize the need for effective monitoring and management strategies to control subclinical mastitis and its impact on herd health and milk production. Further research and implementation of preventive measures are warranted to address this significant health concern in dairy herd.

The primary causative factors frequently encountered are bacteria, with over 150 Gram-positive and Gram-negative bacterial strains identified as mastitis pathogens. These bacteria can be categorized into contagious, transmitted from other infected quarters, and environmental, originating from the surrounding environment (Ashraf and Imran, 2020; Cobirka *et al.*, 2020; Krishnamoorthy *et al.*, 2021; Ndahetuye *et al.*, 2019 and Ruegg, 2017).

The prevalence of antimicrobial resistance and genetic determinants in bacteria causing bovine mastitis is a growing concern in Algeria and worldwide. Several studies have investigated this issue, focusing on pathogens such as *Staphylococcus aureus* (*S. aureus*) and *Escherichia coli* (*E. coli*), Table 2.

Table 2. Antibiotic susceptibility profiles and antimicrobial resistance genes detected in *S. aureus* and *E. coli* isolates from Algerian dairy cows. The data reveal widespread resistance, including methicillin-resistant *S. aureus* (MRSA) and multidrug-resistant (MDR) *E. coli*, with the presence of key resistance genes, highlighting concerns over antibiotic misuse and the need for improved surveillance and control measures.

Agent	Antimicrobial resistance profile	MDR	Antimicrobial resistance genes	References
<i>Escherichia coli</i>	Trimethoprim-Sulfamethoxazole, Chloramphenicol, Gentamicin, Colistin, Amoxicillin–Clavulanic acid, Kanamycin, Enrofloxacin, Cefotaxime, Tetracycline, Ampicillin, Nalidixic acid, Amoxicillin.	92,8%	CTX-M	(Ghallache et al., 2021)
<i>Escherichia coli</i>	Amoxicillin, Tetracycline, Amoxicillin-Clavulanic acid, Trimethoprim-Sulfamethoxazole, Doxycycline, Ciprofloxacin.	38,4%	tetA, blaTEM-1, qnrB, qnrA, aadA1, aac(3)-Ib	(Tahar et al., 2020)
<i>Staphylococcus aureus</i>	Erythromycin, Ciprofloxacin, Penicillin, Tetracycline, Gentamicin, Bacitracin	/	Absence of the mecA gene	(Benhamed and Kihal, 2013)
<i>Staphylococcus aureus</i>	Penicillin, Tetracycline, Kanamycin, Erythromycin, Clindamycin, Fusidic acid	16,4%	Absence of the mecA or mecC genes	(Akkou et al., 2016)
<i>Staphylococci</i>	Penicillin, Oxacillin, Clindamycin, Erythromycin, Cefoxitin, Amoxicillin-Clavulanic acid, Vancomycin, Tetracycline	87,71%	mecA, tetM, blaZ	(Saidi et al., 2015)
<i>Staphylococcus aureus</i>	Penicillin, Tulathromycin, Nalidixic acid, tetracycline, Clindamycin, Erythromycin, Pirlimycin, Tylosin, Tilmicosin, Spectinomycin	100%	blaZ, blaI, blaR, erm B, tet K	(Zaatout et al., 2019)
<i>Staphylococcus aureus</i>	Penicillin, Amoxicillin-Clavulanic acid, Erythromycin, Clindamycin, Tetracycline, Cefoxitin, Trimethoprim-Sulfamethoxazole	30,64%	mecA, blaZ	(Bouzidi et al., 2023)

S. aureus is a resilient bacterium that can survive in various environmental conditions, making it a major cause of intra-mammary infections in dairy cattle. The bacterium is highly prevalent in dairy farms, with its prevalence varying across different regions. The robust nature of *S. aureus* allows it to thrive in various settings, including dairy and beef production, processing, and supply lines. It is also known to be prevalent in raw meat, raw milk, equipment, and personnel who handle milk and meat. This widespread presence of *S. aureus* in the dairy farming environment emphasizes the need for stringent hygiene practices to prevent biofilm formation and the spread of infection within herds (Gizaw et al., 2023).

Akkou *et al.* (2016) reported *S. aureus* isolation in 74% of herds with 5–25 cows, offering valuable insights into mastitis epidemiology in smaller dairy farms. Furthermore, this pathogen was directly implicated in 29.8% of the cases, emphasizing its role as a primary contributor to the mastitis burden. The findings are consistent with other provincial studies in Algeria, such as the investigation conducted in Oran by Benhamed and Kihal (2013), which reported a notably high frequency of *S. aureus* in bovine mastitis. In this specific region, *S. aureus* was identified as the causative agent in 38.9% of both clinical and subclinical mastitis cases. These regional studies contribute to the broader understanding of the distribution and prevalence of *S. aureus* in different areas of the country. However, it is essential to approach the comparison of results from various prevalence studies with caution due to potential differences in sample selection and bacterial cultivation techniques (Akkou *et al.*, 2016).

Several studies have emphasized the necessity of accurately identifying *S. aureus* infected cows to ensure the effective implementation of mastitis control programs (Akkou *et al.*, 2016; Benhamed and Kihal, 2013; Bouzidi *et al.*, 2023 and Zaatout *et al.*, 2019).

Public health is seriously concerned when it comes to bacteria identified in milk that have genes for antibiotic resistance, especially when dairy farming is involved. The abuse or misuse of antibiotics in animal husbandry has the potential to promote the establishment of resistant diseases, especially in developing countries (Sipahi *et al.*, 2023). Antibiotic-resistant bacteria not only threaten the health of diseased animals but also have the potential to impact healthy animals, which makes this issue especially pertinent in the case of bovine mastitis. Antibiotic resistance that makes it difficult to treat mastitis might cause dairy farmers to lose money and become less productive. Promoting appropriate antibiotic usage in the dairy business and increasing public knowledge of the effects of antimicrobial resistance are crucial in order to reduce this growing threat (Dong *et al.*, 2022) and (Sipahi *et al.*, 2023).

The veterinary clinics prolonged and frequent use of antibiotics like ampicillin, amoxicillin, cloxacillin, penicillin G, oxacillin, streptomycin and tetracycline has resulted in the emergence of resistant strains of bacteria (Saidi *et al.*, 2015). Although little is known about the use of vancomycin in dairy cattle, the Saidi *et al.*, 2015 study emphasizes the high frequency of 76.2% of vancomycin-resistant *S. aureus* isolates in dairy animals as well as the lack of *vanA* in study (Saidi *et al.*, 2021). However, no research has been conducted elsewhere that reports the presence of vancomycin-resistant *S. aureus* in bovine mastitis in Algeria.

The most commonly antimicrobial resistance determined was against penicillin, erythromycin and tetracycline, Table 2. The study of Saidi *et al.*, 2015 conducted in Alger found that *S. aureus* isolates from bovine mastitis were highly resistant to penicillin (95.23%), oxacillin (80.95%), clindamycin (80.95%) and erythromycin (76.19%). MRSA strains were also detected, being methicillin-resistant and resistant to multiple antibiotics.

Almost the same results were reported by other authors (Akkou *et al.*, 2016, Benhamed and Kihal, 2013b) especially on resistance to penicillin. The study conducted in Tiaret found that 30.64% of *S. aureus* isolates from subclinical bovine mastitis were multi-drug resistant, and 8% were MRSA isolates. The study also detected virulence genes, including *eta*, *icaA*, *icaD*, *etb*, *luk E-D* and *sea*, which play an important role in the pathogenesis of subclinical bovine mastitis (Bouzidi *et al.*, 2023). Bouzidi's results brought to light the prevalence of several genetic components in MRSA and methicillin-susceptible *S. aureus* (MSSA) isolates that encode resistance to antibiotics. This may be the result of high antibiotic use in dairy cows, which promotes the selection of antibiotic-resistant strains, or it may be the result of mobile genetic elements encoding antibiotic resistance being exchanged (via conjugation, transposition and transduction) from resistant *S. aureus* strains (Bouzidi *et al.*, 2023).

However, there were variations in the frequency of Multi-drug resistance phenotypes and resistance genes found in each study. In the study of Saidi *et al.*, 2015, the distribution of antibiotic resistance genes was as *mecA* (100%) and *tetM* (100%) followed by *blaZ* (42.85%), whereas Bouzidi *et al.* in 2023, found that out of 59 penicillin-resistant isolates, 14 harbored the *blaZ* gene; one of them co-harbored the *mecA* gene. Genes conferring resistance to tetracycline (*tet K*), penicillin (*blaZ*, *bla I* and *bla R*) and macrolide–lincosamide–streptogramin B (*erm B* and *ermA*) were also detected in the *S. aureus* from the study of Zaatout *et al.* in 2019. However, no *mecA* genes were detected in the studies of Akkou *et al.* in 2016 and Benhamed and Kihal in 2013.

The *blaZ* encodes a protein penicillinase which is a type of beta-lactamase. This protein inactivates penicillin by hydrolyzing the beta-lactam ring, and it is responsible for beta-lactam resistance in Staphylococci. Staphylococcal beta-lactamase acts as an extended-spectrum beta-lactamase and contributes to borderline resistance to penicillin (Nomura *et al.*, 2020). The high incidence of penicillin resistance in Staphylococci is largely due to the presence of the *blaZ* gene (Ferreira *et al.*, 2016).

The *mecA* gene is responsible for encoding a penicillin-binding protein in methicillin resistant *Staphylococcus spp.* However, studies have shown that *mecA* may also promote resistance to other beta-lactam antibiotics (Abebe *et al.*, 2020) and (Gueimonde *et al.*, 2013). For this reason, The presence of the *mecA* gene along with the *blaZ* gene in bacteria is very important and poses a threat to public and animal health due to the constant evolution of drug resistance in bacteria (Andersson *et al.*, 2020) and (Tóth *et al.*, 2020). Different prevalences can be detected in studies conducted in different regions or in studies conducted in the same region at different times (Sipahi *et al.*, 2023).

Furthermore, the occurrence of tetracycline-resistant and macrolide/clindamycin-resistant genes among bovine *Staphylococci* isolates has been observed in different countries, suggesting their role in promoting the evolution and development of antibiotic resistance (Mzee *et al.*, 2023), (Saidi *et al.*, 2019 and Zaatout *et al.*, 2019).

Along with *Staphylococci* and *Streptococci*, *E. coli* is one of the most commonly isolated causal organisms linked to bovine intramammary infection (Burvenich *et al.*, 2003). *E. coli* multidrug resistance is a concerning problem that is becoming more and more prevalent in both human and veterinary medicine globally.

Although, *E. coli* is naturally sensitive to practically all clinically important antibiotics, but, it is able to acquire resistance genes, mostly through horizontal gene transfer (Poirel *et al.*, 2018). *E. coli* in milk is a cause for concern due to its association with fecal contamination (Kagkli *et al.*, 2007).

In Algeria, the prevalence of *E. coli* in milk samples was 26.94% in Kolea and Birtouta Province (Ghallache *et al.*, 2021) close to the prevalence 26% found in Medjana province, Sidi Embarak province, and El Achir region (Tahar *et al.*, 2020). Indeed, mastitis frequently involves the bacteria *E. coli* (Goulart and Mellata, 2022). When it comes to the contamination of milk with *E. coli*, farm hygiene is a major factor.

The prevalence of environmental *E. coli* may be associated with poor farm cleanliness and poor slope of stable areas. Feces which are common sources of *E. coli* can contaminate the premises directly or indirectly through bedding, calving stalls, udder wash water, and milker's hands (Abutarbush, 2010)

It is imperative to follow proper hygiene procedures, such as cleaning the udders before milking to minimizing bacterial contaminations (Seferoğlu and Kirkan, 2022). Both human digestive tracts and those of

numerous animal species used in food production contain *E. coli*. The genetic flexibility and adaptability of this bacteria to constantly changing environments allows it to acquire a great number of antimicrobial resistance mechanisms. New strains of multidrug-resistant foodborne bacteria, such *E. coli* that produces extended spectrum beta-lactamase (ESBL), have been linked to the introduction of new types of resistant bacteria in food-producing animals due to the usage of antibiotics that are crucial for human medicine (Ramos *et al.*, 2020).

E. coli in cows with clinical mastitis in 42 different dairy farms have been investigated. The most frequently observed resistance was to amoxicillin 86.5%, tetracycline 75%, amoxicillin-clavulanic acid 59.6%, trimethoprim-sulfamethoxazole 36.5%, doxycycline 13.5%, and ciprofloxacin 13.5%. Multidrug resistance was observed in 38.4% of isolates (Tahar *et al.*, 2020). In a similar study, in the same country for a similar survey, the antibiotic resistance profile of the isolated *E. coli* strains from subclinical mastitis was to amoxicillin 75.3%, nalidixic acid 74.3%, ampicillin 57.7%, tetracycline 52%, cefotaxime 51.5%, enrofloxacin 39.2%, kanamycin 31.9%, amoxicillin/clavulanate 23.3%, colistin 13.4%, gentamicin 12.3%, chloramphenicol 6.2% and 3.1% of the strains were resistant to trimethoprim-sulfamethoxazole. Furthermore, most of the *E. coli* strains (92.8%) were resistant to more than one antibiotic (Ghallache *et al.*, 2021). The extensive usage of amoxicillin, tetracycline, and amoxicillin-clavulanic acid in veterinary medicine, along with their affordability, are the primary causes of the elevated resistance levels seen in the current investigations (Tahar *et al.*, 2020).

Whereas, following the results of these studies the best antibiotics in the fight against mastitis in Algeria are gentamicin and the combination trimethoprim-sulfamethoxazole. The effectiveness of enrofloxacin therapies has been shown in other experimental or clinical investigations (Persson *et al.*, 2015).

Resistance to beta-lactams is possibly the most found in *E. coli* isolates from bovine mastitis (Naranjo-Lucena and Slowey, 2023). Therefore, the observed resistance rate in the studies was quite high indicating that *E. coli* strains producing beta-lactamases were common in Algeria. The blaTEM-1 gene (30.7%) was only present in the ESBL-producing *E. coli* isolates that were detected in the study of Tahar and colleagues in 2020. The bla CTX-M has also been reported in another study from 69.2% of the *E. coli* strains where 78% of the bacteria that were resistant to CTX displayed an ESBL-classic synergy pattern (Ghallache *et al.*, 2021).

The blaTEM-1 and CTX-M genes are significantly present in these isolates, indicating a troubling level of resistance to third-generation cephalosporins. These data further emphasize the frequency of ESBL-producing *E. coli*.

The presence of blaTEM and blaCTX-M types in Enterobacteriaceae isolates from cows is worrisome, as ESBL *E. coli* producers, particularly the CTX-M type, play a significant role in causing human infections, both in healthcare settings and within the community. This raises concerns about the potential transmission and impact on public health (De Angelis *et al.*, 2020).

E. coli resistant to beta-lactams poses a significant concern because these isolates were not only resistant to beta-lactams but also revealed genes encoding resistance to tetracycline, quinolones and aminoglycosides. Furthermore, the simultaneous occurrence of resistance genes for tetracyclines, quinolones and aminoglycosides alongside beta-lactams resistance genes in multidrug-resistant (MDR) *E. coli* suggests a common localization of these genes on mobile genetic elements (Tahar *et al.*, 2020).

Tetracycline is a wide-spectrum bacteriostatic antibiotic used in cattle to treat various infections. However, its misuse has prompted some countries, including European countries, to prohibit its use in animal feed (European Commission, 2019). National studies have shown high rates of tetracycline resistance in *E. coli* with 52% (Ghallache *et al.*, 2021) and 75% (Tahar *et al.*, 2020), which suggests that this issue is widespread with previous reports from Algeria. tetA was the most prevalent tetracycline gene, identified, followed by tetB and tetC genes. Among the isolates resistant to ciprofloxacin, two quinolone genes were detected: qnrB and qnrA.

The examination for genes encoding aminoglycoside-modifying enzymes revealed that the isolates carried the aadA1 and aac(3)-IId genes. Fortunately, none of the tested isolates contained the tetJ, aph(30), aac(60), ant, armA, and aac(60)-Ib cr genes (Tahar *et al.*, 2020).

Resistance to colistin that is of particular importance, have also been detected in 13.4% of the *E. coli* strains (Ghallache *et al.*, 2021). Colistin is currently recognized as the final option for treating infections caused by multidrug-resistant (MDR) Gram-negative bacteria (Mohapatra *et al.*, 2021). It is a widely used antibiotic for enterobacterial infections and growth promotion in veterinary medicine and agriculture. However, the rapid proliferation of mcr resistance genes has been aided by this broad usage. Finding the

prevalence of mcr resistance genes in these reservoirs is crucial for improving the implementation of control and prevention efforts as well as for gaining a more accurate knowledge of the global expansion. Therefore, it would be interesting to look at the plasmid profiles of isolated strains in order to determine the genetic basis of colistin resistance (Kumar *et al.*, 2020).

Conclusion

The study reveals a high prevalence of *S. aureus* and *E. coli* in Algerian dairy cows, both of which exhibit significant antibiotic resistance. Concerns about poor hygiene standards and the misuse of antibiotics on dairy farms are raised by the occurrence of antibiotic-resistant strains and subclinical mastitis in cows. Notably, bacteriological and antimicrobial resistance studies of bovine mastitis have not been fully performed in Algeria, particularly in terms of limited molecular detection of resistance genes. Furthermore, the research suggests that veterinarians and herd managers need to pay close attention to the increasing number of methicillin-resistant *S. aureus* and multidrug-resistant *E. coli* isolates found in Algerian dairy farms. The findings also highlight the importance of developing strategic plans for the treatment, prevention, and control of mastitis in Algeria, as well as the need for further research to evaluate virulence factors and to conduct whole genome sequencing to characterize the virulence and resistance genes of the isolates. The study's results can be used as a baseline for future investigations and to inform treatment policies and antimicrobial strategies.

References

1. Abebe, E; Gugsu, G and Ahmed, M (2020). Review on Major Food-Borne Zoonotic Bacterial Pathogens. J Trop Med 2020, PP : 1-19.
2. Abutarbush, S.M (2010). Veterinary Medicine - A Textbook of the Diseases of Cattle, Horses, Sheep, Pigs and Goats, 10th edition. Can Vet J 51 : 541.
3. Akkou, M ; Antri, K ; Bachtarzi, M.-A ; Bes, M ; Tristan, A ; Dauwalder, O ; Kaidi, R ; Meugnier, H ; Tazir, M and Etienne, J (2016). Phenotypic and genotypic characterization of *Staphylococcus aureus* strains associated with bovine mastitis and nasal carriage of workers in contact to animals in Algeria. Pak. Vet. J., 36 : 184-188.
4. Andersson, D.I ; Balaban, N.Q ; Baquero, F ; Courvalin, P ; Glaser, P ; Gophna, U ; Kishony, R ; Molin, S and Tønjum, T (2020). Antibiotic resistance: turning evolutionary principles into clinical reality. FEMS Microbiology Reviews., 44 :171-188.

5. Ashraf, A ; Imran, M (2020). Causes, types, etiological agents, prevalence, diagnosis, treatment, prevention, effects on human health and future aspects of bovine mastitis. *Anim. Health. Res. Rev.*, 21:36-49.
6. Benhamed, N and Kihal, M (2013). Biodiversity of molecular profile of *Staphylococcus aureus* isolated from bovine mastitis cases in West Algeria. *J. Bacteriol. Res.*, PP : 541-45.
7. Bouzidi, S ; Bourabah, A ; Cheriet, S ; Abbassi, M.S ; Meliani, S and Bouzidi, H (2023). Occurrence of virulence genes and methicillin-resistance in *Staphylococcus aureus* isolates causing subclinical bovine mastitis in Tiaret area, Algeria. *Letters in Applied Microbiology.*, 76 :1-9.
8. Burvenich, C ; Van Merris, V. ; Mehrzad, J ; Diez-Fraile, A ; Duchateau, L (2003). Severity of *E. coli* mastitis is mainly determined by cow factors. *Vet. Res.*, 34 : 521-564.
9. Cobirka, M ; Tancin, V and Slama, P (2020). Epidemiology and Classification of Mastitis. *Animals.*, 10 :2212.
10. Regulation, EU (2019). Commission Regulation (EU) 2019/4 of the European Parliament and of the Council of 11 December 2018 on the manufacture, placing on the market and use of medicated feed, amending Regulation (EC) No 183/2005 of the European Parliament and of the Council and repealing Council Directive 90/167/EEC (Text with EEA relevance). *Off. J. Eur. Union.*, 4 :1-23.
11. De Angelis, G ; Del Giacomo, P ; Posteraro, B ; Sanguinetti, M and Tumbarello, M (2020). Molecular Mechanisms, Epidemiology, and Clinical Importance of β -Lactam Resistance in Enterobacteriaceae. *International Journal of Molecular Sciences.*, 21 : 5090.
12. Dong, L ; Meng, L ; Liu, H ; Wu, H ; Schroyen, M ; Zheng, N and Wang, J (2022). Effect of Cephalosporin Treatment on the Microbiota and Antibiotic Resistance Genes in Feces of Dairy Cows with Clinical Mastitis. *Antibiotics (Basel).*, 11 :117.
13. Ferreira, A.M ; Martins, K.B ; Silva, V.R. da ; Mondelli, A.L and Cunha, M. de L.R. de S. da (2016). Correlation of phenotypic tests with the presence of the blaZ gene for detection of beta-lactamase. *Braz J Microbiol.*, 48 :159-166.
14. Ghallache, L ; Mohamed-Cherif, A ; China, B ; Mebkhou, F ; Boilattabi, N ; Bouchemal, A ; Rebia, A ; Ayachi, A ; Khelef, D and Miroud, K (2021). Antibiotic Resistance Profile of *Escherichia coli* Isolated from Bovine Subclinical Mastitis of Dairy Farms in Algeria from 2017 to 2019. *World's Veterinary Journal.*, 11 :402-415.
15. Gizaw, F ; Kekeba, T ; Teshome, F ; Kebede, M ; Abreham, T ; Berhe, H.H ; Ayana, D ; Edao, B.M ; Waktole, H ; Tufa, T.B ; Abunna, F ; Beyi, A.F ; Abdi, R.D (2023). Multidrug-Resistant *Staphylococcus aureus* Strains Thrive in Dairy and Beef Production, Processing, and Supply Lines in Five Geographical Areas in Ethiopia. *Vet Sci.*, 10 :663.
16. Goulart, D.B and Mellata, M (2022). *Escherichia coli* Mastitis in Dairy Cattle: Etiology, Diagnosis, and Treatment Challenges. *Front. Microbiol.*, 13 : 928346.

17. Gueimonde, M ; Sánchez, B ; G. De Los Reyes-Gavilán, C and Margolles, A (2013). Antibiotic resistance in probiotic bacteria. *Front. Microbiol.*, 4 :202.
18. Kagkli, D.M ; Vancanneyt, M ; Vandamme, P ; Hill, C and Cogan, T.M (2007). Contamination of milk by enterococci and coliforms from bovine faeces. *Journal of Applied Microbiology.*, 103 :1393-1405.
19. Krishnamoorthy, P ; Suresh, K.P ; Jayamma, K.S ; Shome, B.R ; Patil, S.S and Amachawadi, R.G (2021). An Understanding of the Global Status of Major Bacterial Pathogens of Milk Concerning Bovine Mastitis: A Systematic Review and Meta-Analysis (Scientometrics). *Pathogens.*, 10 : 545.
20. Kumar, H ; Chen, B.-H ; Kuca, K ; Nepovimova, E ; Kaushal, A ; Nagraik, R ; Bhatia, S.K ; Dhanjal, D.S ; Kumar, V ; Kumar, A ; Upadhyay, N.K ; Verma, R and Kumar, D (2020). Understanding of Colistin Usage in Food Animals and Available Detection Techniques: A Review. *Animals (Basel).*, 10 :1892.
21. Mohapatra, S.S ; Dwibedy, S.K and Padhy, I (2021). Polymyxins, the last-resort antibiotics: Mode of action, resistance emergence, and potential solutions. *J Biosci.*, 46 :85.
22. Mzee, T ; Kumburu, H ; Kazimoto, T ; Leekitcharoenphon, P ; van Zwetselaar, M ; Masalu, R ; Mlaganile, T ; Sonda, T ; Wadugu, B ; Mushi, I ; Aarestrup, F.M and Matee, M (2023). Molecular Characterization of *Staphylococcus aureus* Isolated from Raw Milk and Humans in Eastern Tanzania: Genetic Diversity and Inter-Host Transmission. *Microorganisms.*, 11 :1505.
23. Naranjo-Lucena, A and Slowey, R (2023). Invited review: Antimicrobial resistance in bovine mastitis pathogens: A review of genetic determinants and prevalence of resistance in European countries. *Journal of Dairy Science.*, 106 : 1-23.
24. Ndahetuye, J.B ; Persson, Y ; Nyman, A.-K ; Tukei, M ; Ongol, M.P and Båge, R (2019). Aetiology and prevalence of subclinical mastitis in dairy herds in peri-urban areas of Kigali in Rwanda. *Trop Anim Health Prod.*, 51:2037-2044.
25. Nomura, R ; Nakaminami, H ; Takasao, K ; Muramatsu, S ; Kato, Y ; Wajima, T and Noguchi, N (2020). A class A β -lactamase produced by borderline oxacillin-resistant *Staphylococcus aureus* hydrolyses oxacillin. *Journal of Global Antimicrobial Resistance.*, 22 :244-247.
26. Persson, Y ; Katholm, J ; Landin, H and Mörk, M.J (2015). Efficacy of enrofloxacin for the treatment of acute clinical mastitis caused by *Escherichia coli* in dairy cows. *Veterinary Record.*, 176 : 673-673.
27. Ramos, S ; Silva, V ; Dapkevicius, M. de L.E ; Caniça, M ; Tejedor-Junco, M.T ; Igrejas, G and Poeta, P (2020). *Escherichia coli* as Commensal and Pathogenic Bacteria among Food-Producing Animals: Health Implications of Extended Spectrum β -Lactamase (ESBL) Production. *Animals (Basel).*, 10 :2239.
28. Rana, E.A ; Fazal, M.A and Alim, M.A (2022). Frequently used therapeutic antimicrobials and their resistance patterns on *Staphylococcus aureus* and *Escherichia coli* in mastitis affected lactating cows. *Int J Vet Sci Med.*, 10 :1-10.
29. Ruegg, P.L (2021). What Is Success? A Narrative Review of Research Evaluating Outcomes of Antibiotics Used for Treatment of Clinical Mastitis. *Front. Vet. Sci.*, 8 :639641.

30. Ruegg, P.L (2017). A 100-Year Review: Mastitis detection, management, and prevention. *Journal of Dairy Science.*, 100 :10381-10397.
31. Saidi, R ; Cantekin, Z ; Khelef, D ; Ergün, Y ; Solmaz, H and Kaidi, R (2015). Antibiotic Susceptibility and Molecular Identification of Antibiotic Resistance Genes of Staphylococci Isolated from Bovine Mastitis in Algeria. *Kafkas Universitesi veteriner fakultesi Dergisi.*, 21.
32. Saidi, R ; Cantekin, Z ; Mimoune, N ; Ergun, Y ; Solmaz, H ; Khelef, D and Kaidi, R (2021). Investigation of the presence of slime production, VanA gene and antiseptic resistance genes in Staphylococci isolated from bovine mastitis in Algeria. *Veterinarska stanica.*, 52 : 0-0.
33. Saidi, R ; Mimoune, N ; Baazizi, R ; Benaissa, M.H ; Khelef, D and Kaidi, R (2019). Antibiotic susceptibility of Staphylococci isolated from bovine mastitis in Algeria. *J Adv Vet Anim Res.*, 6 :231-235.
34. Seferoğlu, Y and Kirkan, Ş (2022). Bovine *Escherichia coli* Mastitis and Effects on Milk Microbiota. *Animal Health Production and Hygiene.*, 11 :56-65.
35. Sipahi, N; Kaya, E ; Çelik, C and Pınar, O (2023). The Characterization and Beta-Lactam Resistance of Staphylococcal Community Recovered from Raw Bovine Milk. *Antibiotics.*, 12 :556.
36. Tahar, S ; Nabil, M.M ; Safia, T ; Ngaiganam, E.P ; Omar, A ; Hafidha, C ; Hanane, Z ; Rolain, J.-M and Diene, S.M (2020). Molecular Characterization of Multidrug-Resistant *Escherichia coli* Isolated from Milk of Dairy Cows with Clinical Mastitis in Algeria. *Journal of Food Protection.*, 83 :2173-2178.
37. Tommasoni, C ; Fiore, E ; Lisuzzo, A and Ganesella, M (2023). Mastitis in Dairy Cattle: On-Farm Diagnostics and Future Perspectives. *Animals (Basel).*, 13 : 2538.
38. Tóth, A.G ; Csabai, I ; Krikó, E ; Tózsér, D ; Maróti, G ; Patai, Á.V ; Makrai, L ; Szita, G and Solymosi, N (2020). Antimicrobial resistance genes in raw milk for human consumption. *Sci Rep.*, 10 : 7464.
39. Zaatout, N ; Ayachi, A ; Kecha, M and Kadlec, K (2019). Identification of staphylococci causing mastitis in dairy cattle from Algeria and characterization of *Staphylococcus aureus*. *J Appl Microbiol.*, 127 : 1305-1314.